

## Contribution to the Evaluation of Antibacterial Activity of *Allium Sativum* L. (Garlic) Essential Oil and Fresh Juice

Zohra Chekroud\*

Research laboratory of Interactions,  
Biodiversity, Ecosystems and Biotechnology,  
University of Skikda,  
Skikda, Algeria

Hanene Silini

University of Skikda,  
Skikda, Algeria

Sara Khalfi

University of Skikda,  
Skikda, Algeria

Youcef Redjem

EPH, Laib Deradji,  
El Harrouche, Skikda, Algeria

**Abstract:** The research aims to evaluate in vitro the antibacterial activity of the essential oil and fresh juice of *Allium sativum* L. (garlic), harvested from the region of El Harrouche, Skikda (Algeria). The antibacterial effect of essential oil and fresh juice was assessed by the Aromatogram method against three Gram-clinical bacterial strains (*Escherichia coli*, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae*) and a Gram + strain (*Staphylococcus aureus*) in addition to two reference strains *Escherichia coli* ATCC25922 and *Staphylococcus aureus* ATCC29213. The minimum inhibitory concentration of the essential oil and fresh juice was determined by the solid dilution method. The results showed that the essential oil had an antibacterial effect only on *Staphylococcus aureus* strains (19 and 18mm), whereas the fresh juice exerted an antibacterial effect against all the tested strains, with inhibition zones of 17, 18,19,18, 16 and 14 mm for *Staphylococcus aureus*, *Staphylococcus aureus* ATCC 29213, *Escherichia coli*, *Escherichia coli* ATCC25922 *Pseudomonas aeruginosa* and *Klebsiella pneumoniae* respectively. Significant statistic differences were recorded between the effects of different juice dilutions ( $p = 0.004$ ) where the tested strains were completely resistant at the dilution 5%. The minimum inhibitory concentration of the essential oil was 640  $\mu\text{g/ml}$  for *Staphylococcus aureus*, while those of fresh juice were 640 $\mu\text{g/ml}$  for *Escherichia coli* strains, 1280 $\mu\text{g/ml}$  for *Staphylococcus aureus* strains and 2560 $\mu\text{g/ml}$  for *Pseudomonas aeruginosa* and *Klebsiella pneumoniae*.

**Keywords:** Antibacterial activity, *allium sativum* L., Aromatogram, essential oil, fresh juice, minimal inhibitory concentration

**Received:** 12 February 2019; **Accepted:** 9 April 2019; **Published:** 18 July 2019

### I. INTRODUCTION

Antibiotics are not totally inoffensive for the organism. In addition to their antimicrobial activity, they may provoke undesirable effects [1]. Antibiotics are also treated by the appearance and spread of multiresistant bacteria [2]. Faced with this problem, the use of herbal medicine is more and more relevant [3]. Phytotherapeu-

tics are extracted from medicinal plants and their active ingredients may relieve symptoms and even cure diseases [4]. *Allium sativum* L. is one of the old medicinal plants used as condiment, aliment or for therapeutic purposes [5]. Garlic has been used for centuries in various societies to combat infectious diseases. Historically, it is believed that Louis Pasteur described the antibacterial effect of

\*Correspondence concerning this article should be addressed to Zohra Chekroud, Research laboratory of Interactions, Biodiversity, Ecosystems and Biotechnology, University of Skikda, Skikda, Algeria. E-mail: [chekroudzohra@yahoo.fr](mailto:chekroudzohra@yahoo.fr)

© 2019 The Author(s). Published by KKG Publications. This is an Open Access article distributed under a [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License](https://creativecommons.org/licenses/by-nc-nd/4.0/).

garlic in 1858 for the first time, although no reference is available [6, 7]. Garlic is characterised by an antiviral, antifungal and antiparasitic activities [8, 9, 10, 11]. It has a wide broad of antibacterial activities against Gram positive and Gram negative bacteria [8]. In order to continue to exploit the medicinal plants growing in Algeria and famous for their antimicrobial phytopharmaceutical properties, we tried to detect and highlight the antibacterial power of garlic harvested from the region of El Harrouch, the state of Skikda (Algeria) but mainly to compare the effect of essential oil and fresh juice.

## II. MATERIAL AND METHOD

### A. Isolation and Identification of Bacterial Strains

Pathogenic bacteria were isolated from clinical samples. Gram negative bacteria were isolated on Hektoen medium while Gram positive bacteria were isolated on Chapman medium. Negative bacteria were identified using API 20E system. Gram positive bacteria were identified using the catalase and coagulase tests. The reference strains *Escherichia coli* ATCC25922 and *Staphylococcus aureus* ATCC29213 were provided by Dr. Becheker Imane.

### B. Extraction of Garlic Essential Oil

Essential oil of garlic was extracted using an hydrodistillator in the laboratory of soil chemistry, Skikda university. 100 grams of dry garlic were firstly macerated one hour in 300 ml of distilled water. The homogenate was then put in the hydrodistillator during 2 to 3 hours. Two phases were obtained, the organic one is the essential oil and the aqueous one is the aromatic water. Anhydride magnesium or sodium sulfate was added to eliminate all water particles. Essential oil was then conserved in small glass bottles at 4°C.

Fresh juice was obtained after grinding 60 g of the garlic bulbs and filtration through a gauze. The filtrate was then centrifuged at 3000g during 20 mn and the supernatant was recovered. The fresh juice was then diluted in order to obtain the following dilutions: 100%, 75%, 50%, 20%, 10% and 5% [12] and conserved at 4°C.

### C. Chemical Composition of Garlic Essential Oil

Chemical composition of garlic essential oil collected from El Harrouch region was previously determined by [13] using a chromatograph coupled to a mass spectrometer (MS) type Shimadzu equipped with a capillary column OV17X .

### D. Determination of Antibacterial Activity of the Garlic Essential Oil and Fresh Juice

Antibacterial activity was determined using the method of discs diffusion in solid medium (Aromatogram). The inoculum was prepared from a 18 hours culture on a Mueller Hinton broth (opacity equivalent to 0.5MacFarland) with a first dilution then a second dilution (1/100) to get 106CFU/ml [14]. Filter paper discs (6mm) impregnated with 15 ul of essential oil were deposited on Muller Hinton medium previously inoculated with 1 ml of the bacterial suspension 106CFU/ml . A disc impregnated with distilled water and a second one impregnated with tween 20, ethylic alcohol and distilled water were used as control discs. For the fresh juice the filter discs were impregnated with 5 ul of each dilution. A supplemented disc impregnated with 5 ul of distilled water was used as control disc. Antibacterial activity was determined according to [15]. A supplemented disc impregnated with 5 ul distilled water was used as control disc. Petri dishes were then incubated at 37C for 24h.

### E. Determination of Minimal Inhibitory Concentration of Essential Oil and Fresh Juice

1) *Preparation of the mother solution:* Mother solutions of 2560 ug/ml are good for most of the sensitivity standard ranges [16]. In a sterile flask containing a volume of distilled water, we added a quantity of essential oil to get the concentration 2560 ug/ml [14]. 2% (v/v) of ethylic alcohol of 95% and 0.05% of Tween 80 were added to the mother solution to get an homogenate solution.

2) *Dilution in solid medium:* The technique of dilution in solid medium was used. 4 ml of the initial solution of each extract were diluted in 2ml of sterile distilled water. Serial dilutions were then prepared from each extract: 1/2, 1/4, 1/8, 1/16, 1/32, 1/64, 1/128, 1/256. 2ml of each dilution were then added to Petri dishes containing 18 ml of Muller Hinton medium and inoculated with 1 ml of the bacterial suspension (106CFU/ml) previously prepared .

### F. Bacterial Sensitivity to Antibiotics

The antibacterial activity of garlic essential oil and fresh juice was compared with the activity of the following antibiotics: IPM, CIP, GEN, CL, DO, FF C, FOX, AMC, TIM, AMX, PF, VA, GEN, E, SP, FC, MY, P, OX, AM, CL, C, CTX, FOX, AZM, IPM, K, TOB, ATM, CN, OF, TIM (Appendix) .

A bacterial inoculum was firstly prepared. Four to five bacterial colonies were put in 5 ml of sterile physiological water (0.9%) and well homogenised to get an opacity of 0.5 Mac Farland equivalent to an optical density of 0.08 to 0.1 at 625 nm. Muller Hinton medium

previously versed in Petri dishes was then inoculated with the prepared inoculums using the swabbing method. The bacterial sensitivity was determined according to CA-SFM [17].

### G. Statistical Analysis

Statistical analysis was done using One Way ANOVA test with Greenhouse Geisser Correction at  $p < 0.05$ . Data were treated using an IBM SPSS software.

## III. RESULTS AND DISCUSSION

### A. Microbiological Analysis

During this study we worked on 4 pathogenic bacterial strains. They were identified using macroscopic ,

microscopic and biochemical characters as *Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* in addition to two reference strains *Escherichia coli* ATCC25922 and *Staphylococcus aureus* ATCC29213.

### B. Characteristics of Garlic Extracts

The organoleptic characteristics of garlic essential oil showed that it is of liquid aspect, pungent odour, and a yellow colour. The yield of essential oil extracted from El Harrouch region garlic was 0.02% whereas that of fresh juice was 2.16% (Table 1).

TABLE 1  
YIELDS OF GARLIC EXTRACTS (ESSENTIAL OIL AND FRESH JUICE)

	Essential Oil	Fresh Juice
Weight of Garlic (G)	100 G	60g
Yield of Extraction (%)	0.02%	2.16%
Extraction Method	Hydrodistillation	Grinding And Centrifugation

Essential oils of garlic are not easily extractable and need extraction under vacuum which is not available. In addition to that this plant is poor in essential oils [18]. The obtained yield was less than that obtained from garlic of Mustaghanem, El Taref and Biskra regions (0.2, 0.3, and 1% respectively) [12, 19, 20]. These variations in garlic essential oil yields are explained by the maturity

degree of garlic bulbs, the interaction with the environment (soil, climate) , the time of harvest and the method of extraction [21].

### C. Evaluation of Garlic Extracts Antibacterial Activity

Essential oil of garlic was only active against *Staphylococcus aureus* strains with inhibition zones of 18mm and 19mm (Table 2).

TABLE 2  
CLASSIFICATION OF THE BACTERIAL STRAINS SENSITIVITY ACCORDING TO MOREIRA ET AL. (2005)

Extract	Essential Oil	Fresh Juice (100%)	Fresh Juice (75%)	Fresh Juice (50%)	Fresh Juice (20%)	Fresh Juice (10%)	Fresh Juice (5%)
Bacterial Strain							
<i>Staphylococcus Aureus</i>	18mm (++)	17mm (++)	15mm (++)	13mm (+)	12mm(+)	6mm (-)	0mm (-)
<i>Staphylococcus Aureus Atcc 29213</i>	19mm (++)	18mm (++)	17mm (++)	15mm (++)	13mm (+)	6mm (-)	0mm (-)
<i>E.Coli</i>	0mm (-)	19mm (++)	18mm (++)	15mm (++)	12mm(+)	0mm (-)	0mm (-)
<i>E.Coli Atcc 25922</i>	0mm (-)	18mm (++)	16mm (++)	15mm (++)	13mm (+)	0mm (-)	0mm (-)
<i>Klebsiella Pneumoniae</i>	0mm (-)	16mm (++)	13mm (+)	12mm (+)	7mm (-)	5mm (-)	0mm (-)
<i>Pseudomonas Aeruginosa</i>	0mm (-)	14mm (++)	12mm (+)	11mm (+)	10mm (-)	7mm (-)	0mm (-)

Not Sensitive (-) : Diameter < 8mm , Sensitive (+) : 9mm < Diameter < 14mm , Very Sensitive (++) : 15mm < Diameter < 19mm

This activity was higher than that obtained by [12, 18, 22] who reported inhibition zones of 15, 8 and 12mm respectively. According to [23, 24, 25, 26] high resistance rates were recorded within Gram negative bacteria in comparison with Gram positive bacteria. This may be due firstly to the action of garlic's allicine on Gram positive bacteria [27] and secondly to the presence of lipopolysaccharidic layer in the wall of Gram negative bacteria [28, 29]. [14] however, reported that 56,6% of *Pseudomonas aeruginosa* strains were sensitive to garlic essential oil of El Harrouch region. On the other hand fresh juice was highly active towards the six stains with inhibition zones of 17, 18, 19, 18, 16 and 14mm for *Escherichia coli*, *Escherichia coli* ATCC 25922, *Staphylococcus aureus*, *Staphylococcus aureus* ATCC 29213, *Klebsiella pneumoniae*, and *Pseudomonas aeruginosa* respectively. Strika and his collaborators [30]

demonstrated that domestic fresh juice was more effective against staphylococci strains (31-29 mm) in comparison with Gram negative bacteria. [31] showed that garlic juice effectively inhibited the growth of both *Pseudomonas sp* and *S. aureus* (16mm for each). Statistical analysis showed significant differences between the effects of the different fresh juice dilutions ( $p = 0.004$ ). Bacteria were totally resistant at the dilution 5%. Essential oil of *Allium sativum* L. did not show an important antibacterial activity in comparison with fresh juice. This may be explained by the loss of volatile components during the extraction and/or storage of essential oil and during the incubation period [32].

#### D. Determination of Minimal Inhibitory Concentration

The minimal inhibitory concentration of essential oil towards the two strains of *Staphylococcus aureus* was 640  $\mu\text{g/ml}$  (Fig 1).

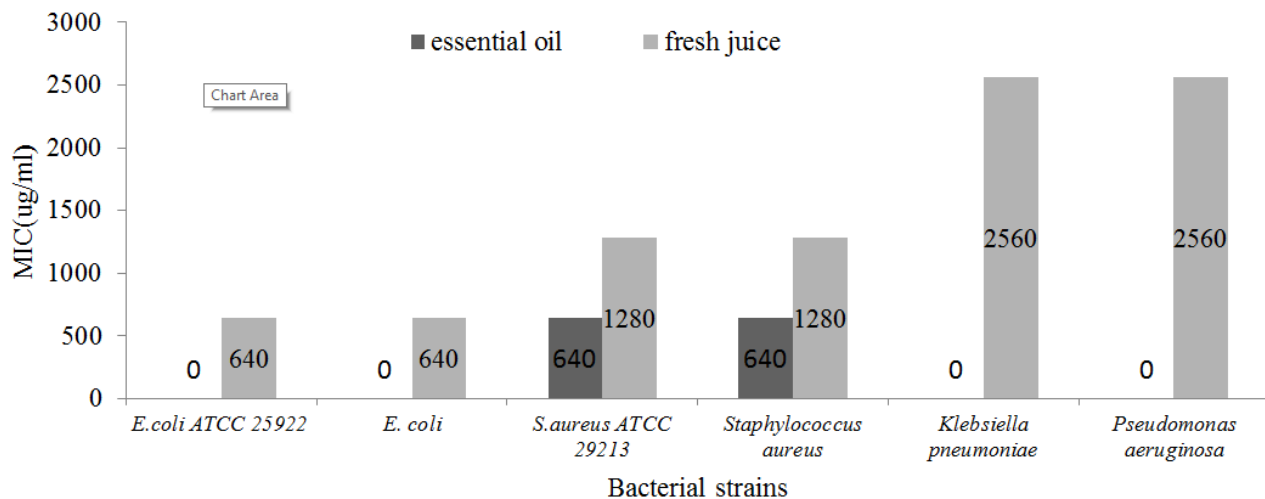


Fig. 1. The minimal inhibitory concentration Of essential oil and fresh juice (Ug/Ml)

The weakest minimal inhibitory concentration of fresh juice was recorded in case of *Escherichia coli* strains (640  $\mu\text{g/ml}$ ) whereas the highest one was recorded with *Klebsiella pneumoniae* and *Pseudomonas aeruginosa* (2560  $\mu\text{g/ml}$ ). [31] reported minimal inhibitory concentrations of 1 and 100 mg/ml for *Pseudomonas aeruginosa* and *Staphylococcus aureus* respectively. The minimal inhibitory concentrations obtained in our study are of high values. This is may be explained by the chemical profile of garlic which is poor in trisulfide diallyl (1.47%) and tetrasulfide diallyl (4.92%) [13]. These two compounds are demonstrated by many authors as the most active compounds in *Allium sativum* L [33, 34].

#### E. Comparison of Antibacterial Activity of Antibiotics and Garlic Extracts

1) *Staphylococcus aureus* ATCC 29213: The reference strain *Staphylococcus aureus* ATCC 29213 is a wild strain (Fig 2). It was very sensitive to the tested antibiotics and very sensitive to essential oil (19mm) and to fresh juice (18mm).

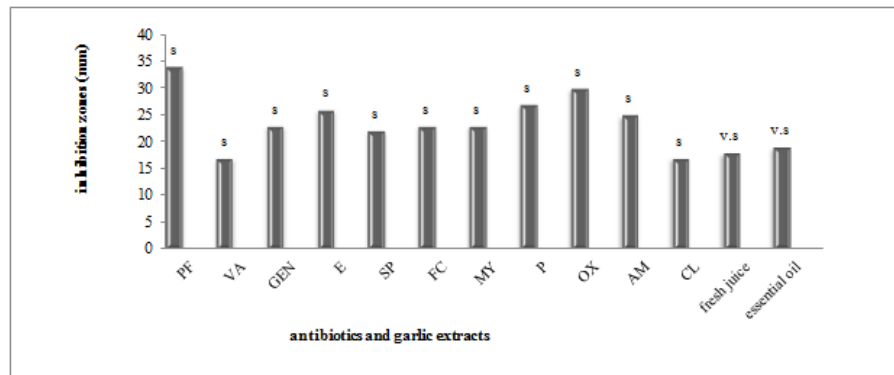


Fig. 2. Comparison between the effect of garlic extracts and antibiotics against *staphylococcus aureus* atcc 29213

2) *Staphylococcus aureus*: *Staphylococcus aureus* was highly sensitive to the garlic extracts, which showed large inhibition zones (18 and 17 mm for the essential oil and

the fresh juice respectively) (Fig 3). Garlic essential oil and fresh juice had better activity than the antibiotics P,AM,OX and CL to which the strain was resistant.

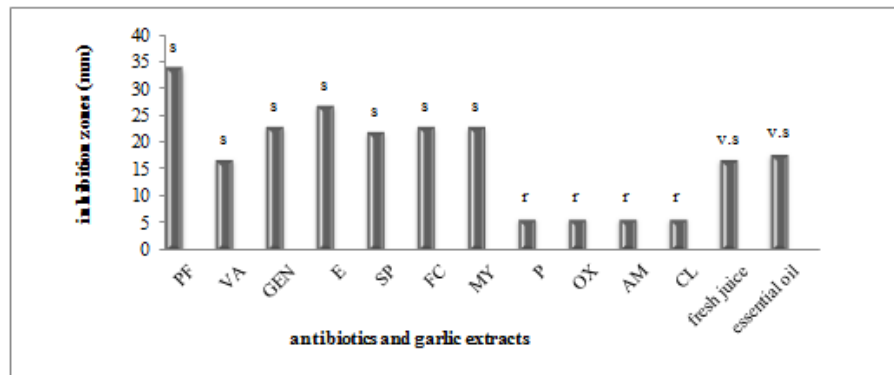


Fig. 3. Comparison between the effect Of garlic extracts and antibiotics against *staphylococcus aureus*

3) *Escherichia coli* ATCC 25922: The reference strain *Escherichia coli* ATCC 25922 which is a wild strain was

sensitive to all the tested antibiotics and highly sensitive to the fresh juice (18mm) (Fig 4).

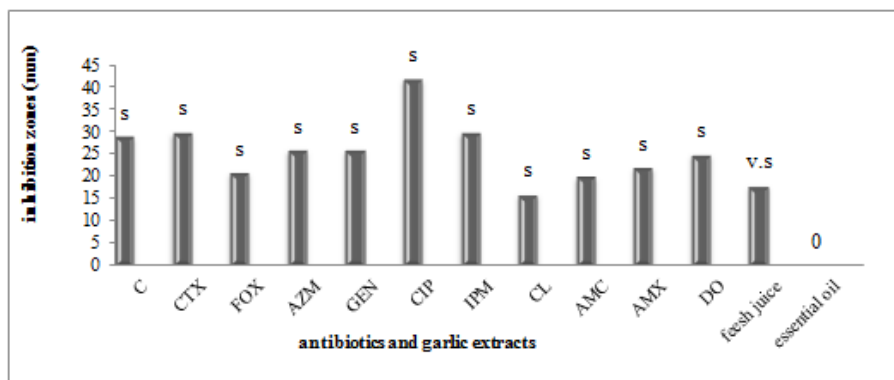


Fig. 4. Comparison between the effect of garlic extracts and antibiotics against *escherichia coli* atcc 25922

4) *Escherichia coli*: *Escherichia coli* has demonstrated high sensitivity toward the fresh juice (17 mm) in com-

parison with the antibiotics AMC,AMX, DO and CL to which it has developed an antibioresistance (Fig 5).

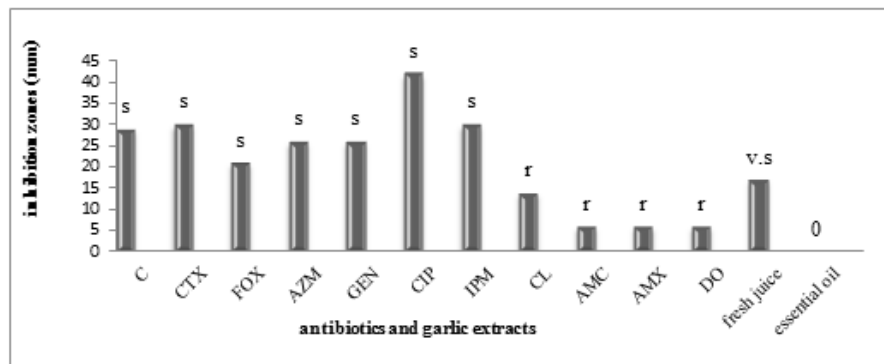


Fig. 5. Comparison between the effect of garlic extracts and antibiotics against *escherichia coli*

5) *Klebsiella pneumoniae*: The obtained results (Fig 6) revealed that *Klebsiella pneumoniae* was very sensitive to fresh juice (16 mm) which had a notorious inhibitory

effect in comparison with the majority of the tested antibiotics TOB, CTX, CL, AMC, CN, OF and TIM whose *Klebsiella pneumoniae* was resistant.

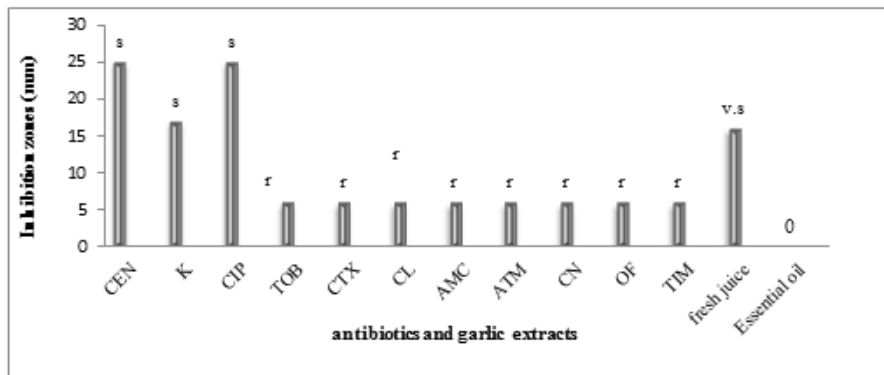


Fig. 6. Comparison between the effect of garlic extracts and antibiotics against *klebsiella pneumoniae*

The advantage of plant extracts is that they don't push the microbes to develop resistance toward them. One of the reasons is that the classical antibiotics are constituted of one active molecule. which makes the microbes easy to synthesis an enzyme able to neutralize or destruct it. For example, the essential oil of *Satureja montana* L. is composed of different molecules, to develop a resistance toward this essential oil, the microbes need to synthesize an enzyme able to degrade all of the active molecules which is impossible or dozen of different enzymes at the same time which is also impossible [35].

#### IV. CONCLUSION

The study of garlic antibacterial activity suggests that this plant is a natural source of chemical molecules that may have important antibacterial activity. This activity differs in function of the extract and the method of extraction which may influence the nature of the extracted components. The fresh juice constitutes an excellent antibacterial product in comparison with essential oil. The obtained results may have possible applications in the

treatment of infectious maladies. Garlic fresh juice and essential oil provide a less expensive source of biomolecules which may be used as complementary drugs with antibiotics, so that they help to face the multiresistant bacteria.

#### ACKNOWLEDGEMENT

The authors gratefully thank the laboratories of bacteriology in El Harrouch and Skikda hospitals; the laboratory of soil chemistry ,department of agronomy , Skikda university; Mme Khadri Sihem and Dr. Becheker Imane, department of sciences of life and of nature, university of august 20th-1955, Skikda.

#### REFERENCES

- [1] P. Berche, J.-L. Gaillard, M. Simonet, and M. Simonet, *Bactériologie: Bactéries Des Infections Humaines*. London, UK: Flammarion médecine-sciences, 1988.
- [2] W. H. Organization, *monographs on selected medicinal plants*. New York, NY: World Health Organization, 1999.

- [3] E. Atefibeibu, "Contribution to the study of tanins and the antibacterial activity of acacia nilotica var adansonii," University of Cheikh Anta Diop, Dakar, Senegal, Unpublished doctoral dissertation, 2002.
- [4] M. Trojan-Rodrigues, T. Alves, G. Soares, and M. Ritter, "Plants used as antidiabetics in popular medicine in Rio Grande do Sul, Southern Brazil," *Journal of Ethnopharmacology*, vol. 139, no. 1, pp. 155–163, 2012. doi: <https://doi.org/10.1016/j.jep.2011.10.034>
- [5] M.-K. Choi, K.-Y. Chae, J.-Y. Lee, and K.-H. Kyung, "Antimicrobial activity of chemical substances derived from S-Alk (en) yl-L-Cysteine Sulfoxide (Alliin) in Garlic, *Allium sativum* L.," *Food Science and Biotechnology*, vol. 16, no. 1, pp. 1–7, 2007.
- [6] L. Bayan, P. H. Koulivand, and A. Gorji, "Garlic: a review of potential therapeutic effects," *Avicenna Journal of Phytomedicine*, vol. 4, no. 1, pp. 1–15, 2014.
- [7] R. A. Hutagalung, Hermawan, S. Magdalena, I. Iskandar, and S. Mastrotillo, "Increasing growth and survival rate of land hermit crabs (*Coenobita* sp.) in artificial habitat through feeding habit," *International Journal of Applied and Physical Sciences*, vol. 3, no. 3, pp. 55–59, 2017. doi: <https://doi.org/10.20469/ijaps.3.50001-3>
- [8] G. Paul and G. Kamel, "Phytothérapie anti-infectieuse," Université de Monastir Springer Verlag France, Paris, Unpublished master's thesis, 2012.
- [9] Y. Yamada and K. Azuma, "Evaluation of the in vitro antifungal activity of allicin," *Antimicrobial Agents and Chemotherapy*, vol. 11, no. 4, pp. 743–749, 1977. doi: <https://doi.org/10.1128/AAC.11.4.743>
- [10] D. Mirelman, D. Monheit, and S. Varon, "Inhibition of growth of *Entamoeba histolytica* by allicin, the active principle of garlic extract (*Allium sativum*)," *Journal of Infectious Diseases*, vol. 156, no. 1, pp. 243–244, 1987. doi: <https://doi.org/10.1093/infdis/156.1.243>
- [11] M. B. Ferlien, B. R. John, E. Sema, P. Silkay, and L. M. Vivien, "Screening of anticholinesterase activity of alkaloids from extracts of ten selected indigenous plants in Mindanao," *Journal of Applied and Physical Sciences*, vol. 4, no. 2, pp. 69–78, 2018. doi: <https://doi.org/10.20474/japs-4.2.4>
- [12] T. Benmeddour, H. Laouar, A. Benabdi, and S. Brahimi, "Evaluation of the activity of antibacterial and antifungal agents in the genus *Allium*," *Courrier du Savoir*, vol. 19, no. 6, pp. 9–14, 2015.
- [13] S. Khadri and N. Boutefnouchet, "Evaluation of the antibacterial activity of essential oil of *Allium sativum* of Eastern Algeria vis-a-vis different strains of *Pseudomonas aeruginosa*," *Journal of Cicbia*, vol. 11, no. 4, pp. 421–428, 2010.
- [14] S. Khadri, "Evaluation of antibacterial activity of garlic (*Allium sativum* L.) essential oil of the Algerian East against different strains of *Pseudomonas aeruginosa*," University of Badji Mokhtar, Annaba, Algeria, Unpublished master's thesis, 2009.
- [15] M. Moreira, A. Ponce, C. Del Valle, and S. Roura, "Inhibitory parameters of essential oils to reduce a foodborne pathogen," *Food Science and Technology*, vol. 38, no. 5, pp. 565–570, 2005. doi: <https://doi.org/10.1016/j.lwt.2004.07.012>
- [16] R. Schwalbe, L. Steele-Moore, and A. C. Goodwin, *Antimicrobial Susceptibility Testing Protocols*. California, CA: CRC Press, 2007.
- [17] Societe Francaise De Microbiologie, "Antibiogram committee of the," 2012. [Online]. Available: <https://bit.ly/3nhZVaZ>
- [18] N. Benzeggouta, "Study of antibacterial activity of infused oils of four medicinal plants known as foods," Mentoury University, Constantine, Algeria, Unpublished master's thesis, 2005.
- [19] N. Benkeblia, "Antimicrobial activity of essential oil extracts of various onions (*Allium cepa*) and garlic (*Allium sativum*)," *Food Science and Technology*, vol. 37, no. 2, pp. 263–268, 2004. doi: <https://doi.org/10.1016/j.lwt.2003.09.001>
- [20] F. Benmeziane, L. Djerroune-Arkoub, A. Damou, K. Hassanand, and H. Zeghad, "Evaluation of antibacterial activity of aqueous extract and essential oil from garlic against some pathogenic bacteria," *International Food Research Journal*, vol. 25, no. 2, pp. 561–564, 2018.
- [21] C. Besombes, "Contribution to the study of hydrothermodynamic extraction phenomenon of aromatic grasses," University of Larochele, Rochelle, France, Unpublished doctoral dissertation, 2008.
- [22] R. Z. Chekki, A. Snoussi, I. Hamrouni, and N. Bouzouita, "Chemical composition, antibacterial and antioxidant activities of tunisian garlic (*Allium sativum*) essential oil and ethanol extract," *Mediterranean Journal of Chemistry*, vol. 3, no. 4, pp. 947–956, 2014. doi: <http://dx.doi.org/10.13171/mjc.3.4.2014.09.07.11>
- [23] K. A. Hammer, C. F. Carson, and T. V. Riley, "Antimicrobial activity of essential oils and other plant extracts," *Journal of Applied Microbiology*, vol. 86,

- no. 6, pp. 985–990, 1999. doi: <https://doi.org/10.1046/j.1365-2672.1999.00780.x>
- [24] E. L. de Souza, N. B. Guerr, T. L. M. Stamford, and E. de Oliveira Lima, “Spices: Alternative sources of antimicrobial compounds to use in food conservation,” *Revista Brasileira de Farmacia*, vol. 87, no. 1, pp. 22–25, 2006.
- [25] D. Elhoussine, B. Zineb, B. Abdellatif et al., “Gc/ms analysis and antibacterial activity of the essential oil of mentha pulegium grown in Morocco,” *Research Journal of Agriculture and Biological Sciences*, vol. 6, no. 3, pp. 191–198, 2010.
- [26] M. Bari, W. Islam, A. Khan, and A. Mandal, “Antibacterial and antifungal activity of solanum torvum (solanaceae),” *International Journal of Agriculture and Biology*, vol. 12, no. 3, pp. 386–390, 2010.
- [27] K. H. Kyung, M. Kim, M. Park, and Y. S. Kim, “Alliinase-independent inhibition of staphylococcus aureus b33 by heated garlic,” *Journal of Food Science*, vol. 67, no. 2, pp. 780–785, 2002. doi: <https://doi.org/10.1111/j.1365-2621.2002.tb10676.x>
- [28] S. Inouye, T. Tsuruoka, K. Uchida, and H. Yamaguchi, “Effect of sealing and tween 80 on the antifungal susceptibility testing of essential oils,” *Microbiology and Immunology*, vol. 45, no. 3, pp. 201–208, 2001. doi: <https://doi.org/10.1111/j.1348-0421.2001.tb02608.x>
- [29] C. Bagamboula, M. Uyttendaele, and J. Debevere, “Inhibitory effect of thyme and basil essential oils, carvacrol, thymol, estragol, linalool and p-cymene towards shigella sonnei and s. flexneri,” *Food Microbiology*, vol. 21, no. 1, pp. 33–42, 2004. doi: [https://doi.org/10.1016/S0740-0020\(03\)00046-7](https://doi.org/10.1016/S0740-0020(03)00046-7)
- [30] I. Strika, A. Basic, and N. Halilovic, “Antimicrobial effects of garlic (allium sativum l.),” *Bulletin of the Chemists and Technologists of Bosnia and Herzegovina*, vol. 47, no. 7, pp. 17–22, 2017.
- [31] V. Kumur and J. John, “Evaluation of antimicrobial effect of allium sativum extract and gomutra,” *World Journal of Pharmaceutical Research*, vol. 7, no. 4, pp. 877–899, 2018.
- [32] F. Chemloul, “Study of antibacterial activity of lavender officinalis essential oil of the region of tlemcen,” University of Abou Bekre Belkaide Tlemcen, Tlemcen, Algeria, Unpublished master’s thesis, 2014.
- [33] E. A. O’Gara, D. J. Hill, and D. J. Maslin, “Activities of garlic oil, garlic powder, and their diallyl constituents against helicobacter pylori,” *Applied and Environmental Microbiology*, vol. 66, no. 5, pp. 2269–2273, 2000. doi: <https://doi.org/10.1128/AEM.66.5.2269-2273.2000>
- [34] S.-m. Tsao and M.-c. Yin, “In vitro activity of garlic oil and four diallyl sulphides against antibiotic-resistant pseudomonas aeruginosa and Klebsiella pneumoniae,” *Journal of Antimicrobial Chemotherapy*, vol. 47, no. 5, pp. 665–670, 2001. doi: <https://doi.org/10.1093/jac/47.5.665>
- [35] V. Christopher, *Natural Alternatives To Antibiotics: Fight Infections By Strengthening The Field*. Jouvence Publisher, 2014.



**Appendix****APPENDIX 1: LIST OF THE TESTED ANTIBIOTICS**

Code	Antibiotic
Am	Ampicillin
Amc	Amoxicillin+Clavulanic Acid
Amx	Amoxicillin
Atm	Aztreonam
Azm	Azithromycin
C	Chloramphenicol
Cip	Ciprofloxacin
Cl	Colistin
Ctx	Cefotaxim
Do	Doxycyclin
E	Erythromycin
Fc	Fusidic Acid
Ff	Fosfomycin
Fox	Cefoxitin
Imp	Imipenem
Gen	Gentamicin
K	Kanamycin
Of	Ofloxacin
My	Lyncomycin
Ox	Oxacillin
P	Penicillin
Pf	Pefloxacin
Sp	Spiramycin
Tim	Ticarcilin
Tob	Tobramycin
V	Vancomycin